

3. Number the bloods in the cards so that the masking tape does not cover the numbers. If you have a “no sample”, just put NS in that number spot as shown in Diagram 1.
4. Tape top and bottom of card. A piece of plain paper under masking tape is really helpful when removing bloods from cards and preventing the tubes of blood from sticking to the tape.
5. Please do NOT use transparent (eg. Scotch) tape.
6. If you are collecting a large number of bloods and you are not sending them in for a couple of days, be sure to refrigerate them after centrifuging until you send them to Lab Services.

**Ensure that you follow proper packaging procedures, as indicated above, so that all samples will be accepted by the Laboratory.**

**Laboratory Services is often working with more than one ranch, so ensure you identify your samples to eliminate any potential identification problems.**

For more information, please contact:

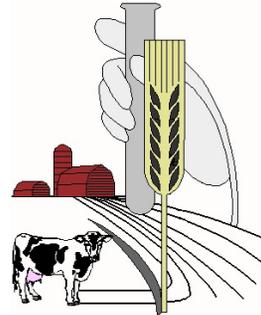
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**Agriculture**

**Agriculture & Food Operations  
Laboratory Services**

## **Procedure for the Collection of Mink Blood for Aleutian Disease Antibody Testing**

**Please call the laboratory to make an appointment for testing.**

The submission of proper samples prevents delays in testing.

Fibrin clots may form even when heparinized capillary tubes are used. Fibrin clots may affect results if the clots cause an insufficient sample for testing.

The following collection techniques may help ensure that your samples are received at the laboratory in good condition. By following the steps marked in red (\*), the occurrence of fibrin clots should be prevented.

## Collection Procedures

1. **On the day of sample collection, do not feed the mink that are to be sampled until after blood has been taken. Water should NOT be withheld. \***
2. **Keep sample (capillary) tubes dry and warm during the process. \***
3. Before clipping the toenail of each mink, dip the clippers in disinfectant and wipe them on a clean towel or cloth.

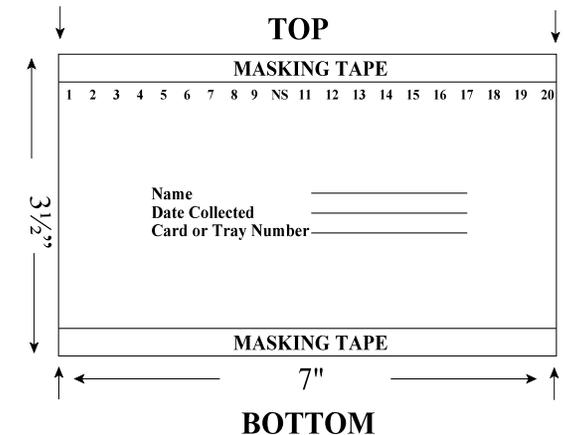
4. Clip the toenail slightly above the vein line (pink area).
5. Fill sample (capillary) tube 3/4 full with mink blood.
6. Plug one end with “critoseal”.
7. **Let the collected blood sit at room temperature (ie. 18-25° C) for at least 30 minutes prior to placing in centrifuge. \***
8. **Place centrifuged sample tubes in cards with no more than 20 tubes per card. \***
9. **Ensure that all bloods remain in an upright position after centrifuging until they are tested by the laboratory. \***
10. Use only sample capillary tubes (Heparinized Micro-Hematocrit) purchased from Laboratory Services/Agricultural office in the past 12 months. Store in a warm, dry area (ie. 18-25° C) and away from sunlight.

**If you have followed the collection procedures and still encounter problems, please contact Laboratory Services to discuss an appropriate course of action.**

## Proper Packaging Procedures

1. Please ensure all cards are approximately 3½" x 7".
2. Label each card as per Diagram 1.

Diagram 1



**Cut tape off at edge of cardboard (top & bottom). There is no need to tape sides of cards or wrap tape over edges.**